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3 **1 Article Type: Perspective**  
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7 **3 Recognizing taste: coding patterns along the neural axis in mammals**  
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3 **22 Abstract**  
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5 23 The gustatory system encodes information about chemical identity, nutritional value, and concentration  
6 24 of sensory stimuli before transmitting the signal from taste buds to central neurons that process and  
7 25 transform the signal. Deciphering the coding logic for taste quality requires examining responses at  
8 26 each level along the neural axis - from peripheral sensory organs to gustatory cortex. From the earliest  
9 27 single fiber recordings, it was clear that some afferent neurons respond uniquely, others to stimuli of  
10 28 multiple qualities. There is frequently a “best stimulus” for a given neuron, leading to the suggestion  
11 29 that taste exhibits “labeled line coding”. In the extreme, a strict “labeled line” requires neurons and  
12 30 pathways dedicated to single qualities (e.g. sweet, bitter, etc.). At the other end of the spectrum,  
13 31 “across-fiber”, “combinatorial”, or “ensemble” coding requires minimal specific information to be  
14 32 imparted by a single neuron. Instead, taste quality information is encoded by simultaneous activity in  
15 33 ensembles of afferent fibers. Further, “temporal coding” models have proposed that certain features of  
16 34 taste quality may be embedded in the cadence of impulse activity. Taste receptor proteins are often  
17 35 expressed in non-overlapping sets of cells in taste buds apparently supporting “labeled lines”. Yet,  
18 36 taste buds include both narrowly- and broadly-tuned cells. As gustatory signals proceed to the  
19 37 hindbrain and on to higher centers, coding become more distributed, and temporal patterns of activity  
20 38 become important. Here, we present the conundrum of taste coding in the light of current  
21 39 electrophysiological and imaging techniques at several levels of the gustatory processing pathway.  
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34 41 **Keywords:** gustatory coding, taste quality, taste bud, geniculate ganglion, nucleus of solitary tract,  
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## 45 Introduction

46 All sensory systems must address the problem of conveying information about the quality, intensity,  
47 and location of sensory stimulation from peripheral receptors to the brain. For both olfaction and taste,  
48 stimuli can be chemically diverse. The olfactory system is known to encode this chemical diversity, in  
49 part, through the use of hundreds of molecular receptors with overlapping receptive ranges. Olfactory  
50 signals from peripheral neurons are carried on circuits that exhibit convergence and distributed  
51 patterns at different stages along the neural axis to encode odor recognition and discrimination  
52 (Laurent 2002; Nara *et al.* 2011; Nunez-Parra *et al.* 2014; Srinivasan and Stevens 2018). The  
53 gustatory system, which serves to detect nutrients, minerals, and toxins, also identifies diverse  
54 chemical structures across broad concentration ranges. The logic of how the mammalian gustatory  
55 system encodes information on chemical identity, i.e. quality coding, is the subject of active  
56 investigation using a variety of experimental approaches and resulting in competing models of taste  
57 coding. The present review examines some of the evidence, interpretations and controversies  
58 regarding gustatory quality coding.

59 Most research on taste quality coding focuses on discriminating “sweet” (for example sugars), “salty”  
60 ( $\text{Na}^+$  salts), “sour” (acids) and so forth. Labeled line coding posits that quality-specific taste receptor  
61 cells (TRCs) (for example “sweet”-specific) synapse only with primary sensory afferent(s) that are  
62 dedicated to that same quality. This, then, establishes a dedicated transmission line from the taste bud  
63 cell to the brain that is “labeled” for a single quality. According to this coding, the different transmission  
64 lines (“sweet”, “salty”, “sour”, etc.) are separate, distinct, and parallel. The sensory afferent neurons  
65 are all highly tuned to transmit one given quality. They are all “specialists” for a given quality.

66 In contrast, combinatorial coding allows more flexibility in the responses of primary afferent fibers.  
67 Thus, a given taste compound can elicit impulses in in an ensemble of several primary afferent fibers,  
68 each of which varies in their response profiles. That is, some fibers might be “sweet-best”, others  
69 might be “salt-best”; they respond robustly to sugars or  $\text{Na}^+$  salts, respectively, while retaining weaker  
70 responses to other tastes (“specialists”). Other fibers in the ensemble may respond quite broadly to  
71 many different taste compounds with no strong preference (“generalists”). However, when activated by  
72 a specific taste compound, the entire ensemble of afferent fibers generates a particular combinatorial  
73 signal that identifies that stimulus. Collectively, the combination of specialists and generalists, not any  
74 individual sensory afferent axon on its own, transmits the information about taste quality. Temporal  
75 coding conveys information in the pattern of impulses in individual primary sensory afferents. Different  
76 taste stimuli may elicit different patterns of action potentials in afferent fibers, which might lead to  
77 differential excitation/inhibition of neurons in the CNS.

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3 78 For theorists, both models present a dilemma: how do multi-sensitive cells convey an unambiguous  
4 79 message that identifies taste quality? The labeled line and across-neuron pattern theories share the  
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6 80 notion that spikes are integrated over time, and ignore the dynamics of firing rate changes that occur  
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8 81 during a taste response. These dynamic aspects of the response may also carry taste information, a  
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10 82 form of signaling called “temporal coding”.

11 83 The origins of labeled line coding in the sensory nervous system might be said to come from René  
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13 84 Descartes, who, in his classical drawing of the innocent cherub toasting his toes (Descartes 1664, p.  
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15 85 27), clearly outlined a labeled line (here, for painful heat) from peripheral sensory organ to the brain  
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17 86 (Roper 2014). However, the first explicit statements of labeled line coding were by Sir Charles Bell  
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19 87 (1811; see Bell and Shaw 1868), and Johannes Müller (1835), who coined the concept, Law of  
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21 88 Specific Nerve Energies (LOSNE), according to which “each type of sensory nerve ending, however  
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23 89 stimulated (electrically, mechanically, etc.), gives rise to its own specific sensation; moreover, each  
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25 90 type of sensation depends not upon any special character of the different nerves but upon the part of  
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27 91 the brain in which their fibers terminate” (Müller 1836). Since then, it has become clear that each  
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29 92 modality is indeed “labeled” insofar as touch, temperature, taste, olfaction, vision, hearing, and so forth  
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31 93 are each transmitted along separate neural pathways. The question, now, is whether such “labeling”  
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33 94 extends to different qualities *within* a sensory modality, such as red versus green color, rose versus  
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35 95 geranium scent, or sweet versus salty taste. That is the crux of the current debate. In certain sensory  
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37 96 systems, such as vision and olfaction, the answer is clearly “no”; colors and odors unarguably display  
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39 97 combinatorial quality coding.

35 98 In this review, we examine the evidence, primarily derived from electrophysiological and imaging  
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37 99 studies at different levels of the taste system, of the responses of receptors and neurons to stimuli  
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39 100 representing different taste qualities. We discuss what the responses at each level suggest about the  
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41 101 logic of coding taste quality.

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### 44 103 **The detectors: coding taste quality in taste bud cells**

46 104 A strict peripheral labeled line coding for taste qualities (sweet, salty, sour, etc.) has been strongly  
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48 105 promoted by some researchers (Barretto *et al.* 2015; Chen *et al.* 2011b; Yarmolinsky *et al.* 2009). The  
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50 106 strongest evidence for such a hard-wired logic for taste quality coding comes from the observation that  
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52 107 taste bud cells express primarily or only one type of taste receptor. Some cells express a few to  
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54 108 several members of the Tas2R family of receptors which are activated by bitter-tasting compounds  
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56 109 (Behrens *et al.* 2007; Mueller *et al.* 2005). Other TRCs may express heterodimeric Tas1R family  
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58 110 receptors, which are activated by either sweet- or umami-tasting compounds (Dando *et al.* 2012;

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3 111 Nelson *et al.* 2002; Nelson *et al.* 2001). Yet other cells are dedicated for sour taste sensing (Huang *et al.* 2006). However, some fraction of taste cells do express taste receptors for more than one quality  
4 112 (Dando *et al.* 2012). The relatively non-overlapping pattern of receptor expression led to the proposal  
5 113 that, similar to insects, mammals use a hard-wired logic for coding taste quality (Yarmolinsky *et al.*  
6 114 2009). That is for example, Tas2R-expressing TRCs, when stimulated, activate a dedicated subset of  
7 115 afferent fibers which would encode the bitter taste quality. Other dedicated TRCs and nerve fibers  
8 116 would convey sweet and so on. This taste quality-dedicated TRCs constitute the beginning of a  
9 117 labeled line for “bitter” or “sweet”, respectively, that is maintained along the taste axis to the gustatory  
10 118 cortex.  
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12 120 The question is how well do the responses of individual taste bud cells mirror the seemingly  
13 121 compartmentalized, non-overlapping pattern of expression of the various taste receptors. The taste  
14 122 quality sensitivity and selectivity of specific populations of taste bud cells have been examined through  
15 123 both, electrophysiological and Ca<sup>2+</sup> imaging methods (Tomchik *et al.* 2007; Yoshida *et al.* 2009;  
16 124 Yoshida *et al.* 2018), using several distinct *ex vivo* preparations. Using the combination of  
17 125 transgenically identified taste bud cell types and apical stimulation with a variety of taste stimuli, the  
18 126 response profiles of taste bud cell types have been studied electrophysiologically (Yoshida *et al.* 2009)  
19 127 and via Ca<sup>2+</sup> imaging (Caicedo *et al.* 2002; Tomchik *et al.* 2007). Very consistently, Type II cells  
20 128 respond best to sweet, bitter or umami taste stimuli. “Bitter-best” taste cells are the most narrowly  
21 129 tuned and respond almost exclusively to bitter compounds (Yoshida *et al.* 2009b). In contrast, some  
22 130 “sweet-best” TRCs are more broadly tuned such that, in addition to sucrose, some also respond to salt  
23 131 (NaCl) and/or umami stimuli (monosodium glutamate, MSG). Type III cells from fungiform taste buds  
24 132 consistently respond to acid (sour) stimuli, and each cell typically responds to multiple acids (citric,  
25 133 acetic or HCl). Thus, tuning, measured in the electrical responsivity of cells from fungiform taste buds  
26 134 (Yoshida *et al.* 2009), is generally similar to that measured by the Ca<sup>2+</sup> responses of Type II cells from  
27 135 mouse circumvallate taste buds (Tomchik *et al.* 2007). Further, in both studies, responses to acids  
28 136 were limited to Type III cells.

29 137 Type III cells in mouse fungiform papillae fell into 2 groups with. approximately 75% responding only to  
30 138 acids, the rest being broadly tuned, with responses to salty, umami, and/or bitter stimuli in addition to  
31 139 acids. This observation differed conspicuously the Ca<sup>2+</sup> imaging study which reported that all or most  
32 140 Type III cells in mouse circumvallate taste buds were both sour-responsive and broadly tuned  
33 141 (Tomchik *et al.* 2007). Whether these differences are attributable to differences in methodology or in  
34 142 the taste bud fields examined (fungiform vs. circumvallate) remains to be determined.

35 143 Another question that has been explored electrophysiologically in mouse fungiform taste bud cells is  
36 144 how diverse stimuli that produce similar taste perception are represented in the initial receptor cells.

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3 145 For example, many sugars (sucrose, fructose, etc.), artificial sweeteners (saccharin, sucralose, etc.)  
4 146 and certain proteins (Monellin, Thaumatin, Brazzein, etc.) all elicit sweet taste. Similarly, there are  
5 147 numerous chemically diverse compounds, all of which elicit bitter taste. To test whether TRCs respond  
6 148 identically to diverse stimuli of a given quality (for example “bitter”) or can discriminate among  
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8 149 **perceptually similar compounds**, responses were recorded to a battery of bitter-tasting compounds  
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10 150 (Yoshida *et al.* 2018). Type II TRCs from fungiform and circumvallate taste buds showed considerable  
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12 151 heterogeneity in their responses to this battery of bitter chemicals. **Some bitter stimuli elicited**  
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14 152 **responses in 5-8 times as many taste cells as did other bitter compounds. That is, taste compounds**  
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16 153 **that are perceived as having similar taste may produce very different patterns of activation among**  
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18 154 **taste bud cells**

19 155 Yoshida *et al.* (2018) also demonstrated that bitter-sensitive cells as a population displayed  
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21 156 **considerable heterogeneity. When tested with** 10 bitter compounds, some were selective for only **a**  
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23 157 **single stimulus while** others responded broadly to as many as 9 of the 10 stimuli tested. Such  
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25 158 heterogeneous responses among bitter-sensitive taste cells had also been demonstrated using  
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27 159 functional imaging of rat and mouse circumvallate taste bud cells (Caicedo *et al.* 2002; Caicedo and  
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29 160 Roper 2001). The family of bitter taste receptors Tas2Rs, includes ≈35 diverse members and each of  
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31 161 these Tas2Rs is activated by a different complement of bitter compounds (Lossow *et al.* 2016). In  
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33 162 both, human and mouse, some Tas2rs are narrowly tuned and others that can be activated by large  
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35 163 numbers of bitter tasting compounds (Lossow *et al.* 2016; Meyerhof *et al.* 2010). Thus, the selectivity  
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37 164 of bitter sensitive TRCs would be defined by the expression of different combinations of Tas2Rs.

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39 165 **All molecular receptors for bitter tastants, Tas2Rs, were reported to be co-expressed in some TRCs**  
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41 166 **with the interpretation that discrimination among bitter stimuli could not occur (Adler *et al.* 2000),** More  
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43 167 comprehensive analyses showed that only limited numbers of Tas2Rs are expressed per TRC, and  
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45 168 in various combinations (Behrens *et al.* 2007; Matsunami *et al.* 2000). **The electrophysiological and**  
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47 169 **Ca<sup>2+</sup> imaging results above also demonstrate that the initial hypothesis (Mueller *et al.* 2005) for how**  
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49 170 **bitter taste quality is coded in the periphery was likely incorrect. Combinatorial** expression of Tas2Rs  
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51 171 in individual TRCs could, in principle, **form a basis for** discriminating among different bitter compounds,  
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53 172 but it is unclear whether such discrimination exists along the taste neural axis or even behaviorally.

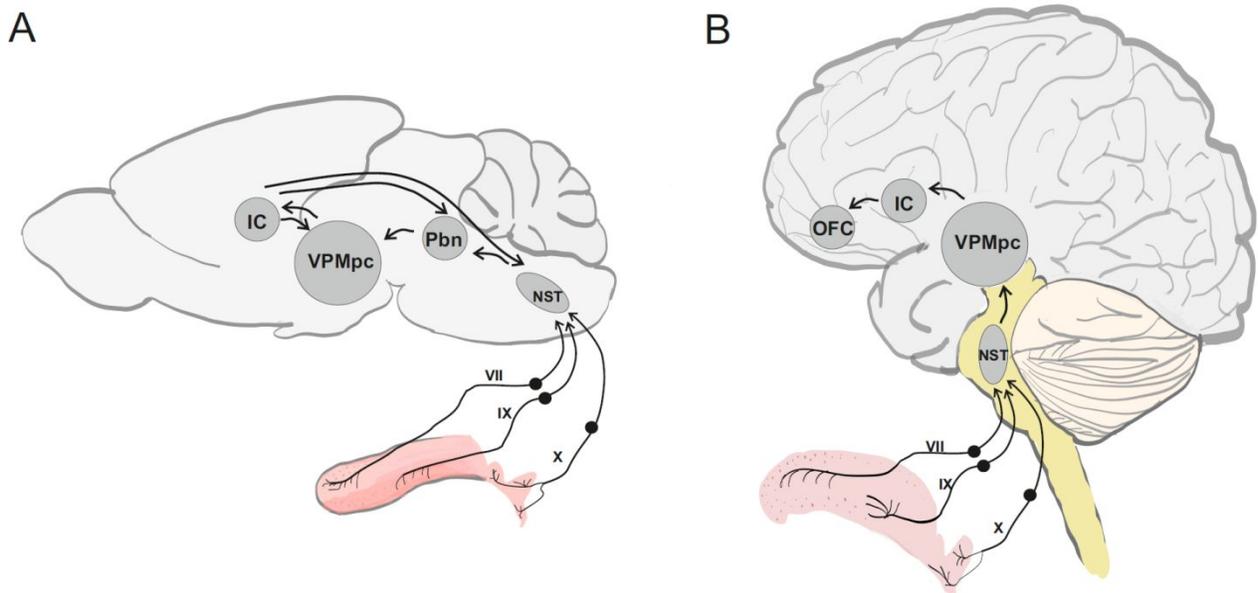
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55 173 Taken together, electrophysiological and Ca<sup>2+</sup> imaging data indicate that taste buds contain many  
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57 174 taste receptor cells dedicated to detect one of 5 basic taste qualities. These may provide the basis for  
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59 175 discrimination across basic taste qualities. However, taste buds also contain TRCs that respond to  
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61 176 multiple taste qualities (Caicedo *et al.* 2002; Tomchik *et al.* 2007; Yoshida *et al.* 2009). **These**  
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63 177 **multiply-responsive** cells may reflect information processing (divergence and convergence of signals)  
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65 178 that occurs within taste buds via cell-cell synaptic interactions (Chaudhari 2014; Dando and Roper

2009; Huang *et al.* 2009; Huang *et al.* 2007). Moreover, some taste cells express multiple types of taste receptors. For instance, a subset of taste cells expresses all three T1R subunits and responds to both sweet and umami compounds (Dando *et al.* 2012; Kushihara *et al.* 2013). Whether broadly tuned TRCs serve a distinct role from narrowly tuned TRCs as well as the contribution of broadly-tuned TRCs to coding of taste signals remain, however, still unclear.

Taste quality coding begins with the sensitivities of individual receptor cells within taste buds. The synaptic connections between these cells and gustatory nerve fibers is a major unknown at present. Understanding convergence or divergence at these peripheral synapses will be key to understanding the initial coding of taste signals in the periphery.

### Quality coding in the first neurons of the taste pathway

How do primary sensory afferent neurons transmit taste information to the central nervous system (CNS; see Figure 1) and how does activity in primary afferents represent taste quality (sweet, salty, sour, etc.)?



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194 **Figure 1.** Schematics of the rodent (A) and human (B) gustatory pathways with a focus on peripheral and  
195 thalamo-cortical relays. In both species, information is conveyed via cranial nerves VII, IX, and X from the tongue  
196 to the brainstem. NST: nucleus of the solitary tract, Pbn: parabrachial nucleus, VPMpc: parvocellular portion of  
197 the ventroposteromedial nucleus of the thalamus, IC: insular cortex, OFC: orbitofrontal cortex.

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3 199 Electrophysiological recordings and Ca<sup>2+</sup> imaging studies from primary sensory afferent neurons  
4 200 (single fibers or ganglion neuron somata) have been carried by several groups. Some form of  
5 201 combinatorial coding in taste was originally suggested by Pfaffmann (1941) based on early  
6 202 electrophysiological recordings from afferent fibers that innervated taste buds in the cat. Single units  
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8 203 were found that responded to lingual stimulation with more than one taste compound (for example  
9 204 quinine or HCl or both). That many fibers were not limited to excitation by a single taste quality, was  
10 205 inconsistent with a labeled line coding scheme. This led Pfaffmann (ibid) to conclude “[...] sensory  
11 206 quality does not depend simply on the “all or nothing” activation of some particular fiber group alone,  
12 207 but on the pattern of other fibers active.” Other investigators elaborated and extended this model to  
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14 208 encompass the widespread co-activation of a large number of sensory afferent fibers, with different  
15 209 combinations of the same fibers constituting the code for different taste qualities. This was termed  
16 210 “cross-fiber coding” and was held as the polar opposite of labeled line coding (Erickson 2008).  
17 211 According to cross-fiber coding, activity in any single fiber on its own does not convey information  
18 212 about sweet, sour, salty, etc. Only the combined activity of many fibers generates the code. Some  
19 213 resolution of these two opposite concepts—labeled line versus combinatorial coding—was obtained by  
20 214 Frank and Pfaffmann (1969). They recorded from single sensory afferent fibers from the tongues of  
21 215 hamsters and observed that although many fibers did indeed respond to multiple taste stimuli, the  
22 216 most effective stimulus of a fiber was predictive of the relative effectiveness of the other stimuli. These  
23 217 observations suggested that there were fiber “types” organized according to the stimulus that evoked  
24 218 the “best” response. They termed these “sweet-best”, “salt-best” etc. fibers. Although this has been  
25 219 interpreted as a form of labeled line coding, the fact is that activity in a single fiber could not  
26 220 unambiguously distinguish between (strong) excitation by the “best” stimulus versus (weak) excitation  
27 221 by other, less effective stimuli.

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40 222 The observation of “best stimulus” for individual taste afferent fibers has been widely replicated in  
41 223 different laboratories, and in mammalian species ranging from mice to monkeys (Danilova *et al.* 1999;  
42 224 Hellekant and Ninomiya 1994; Sato *et al.* 1975; Tonosaki and Beidler 1989). A further refinement of  
43 225 the distinctions between taste afferents was the recognition that some neurons respond principally or  
44 226 exclusively to one stimulus type, usually sugars – the so-called “specialist” neurons; other neurons  
45 227 responded to a variety of electrolytes that might produce sour, bitter or salty tastes (reviewed by Frank  
46 228 *et al.* 2008). Specialist and generalist neurons have been detected electrophysiologically as single-  
47 229 fiber recordings on afferent nerves and by extracellular recordings in geniculate ganglia. A method  
48 230 applied more recently is functional imaging of sensory afferent neuron activity using genetically  
49 231 encoded Ca<sup>2+</sup> indicators such as GCaMP. Barretto *et al.* (2015) and Wu *et al.* (2015) carried out  
50 232 functional imaging on geniculate ganglion neurons in the mouse and cataloged responses to a battery  
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3 233 of different taste stimuli presented at different concentrations. Those studies verified that about half the  
4 234 ganglion neurons were “specialists” that responded best (and some solely) to a single taste  
5 235 compound, such as sucrose. Specialist neurons could be detected for each of the five “basic” taste  
6 236 qualities (sweet, sour, salty, bitter, umami). The geniculate ganglion also had “generalist” sensory  
7 237 neurons that responded much more broadly to taste stimuli, mirroring the electrophysiological  
8 238 recordings from the primary afferent axons (above).

9 239 The relative proportion of specialist and generalist neurons varied strongly depending on the  
10 240 concentrations of stimuli tested (Wu *et al.* 2015). Importantly, neurons that displayed a specialist  
11 241 profile with a low concentration stimulus were transformed to generalists when the same stimuli were  
12 242 tested at higher concentrations. At concentrations that produced maximal responses, half the neurons  
13 243 exhibited responses to multiple distinct stimuli. Unless half the information from the periphery is  
14 244 discarded, which seems unlikely, a resolution to the question of taste coding is that a cross-fiber code  
15 245 involving a combination of primary afferent axons that vary in their “tuning”, from specialists to  
16 246 generalists, encode taste.

17 247 In addition to the encoding the basic taste qualities, there is a question of how stimuli which produce a  
18 248 similar quality may be discriminated from one another. For instance, in primates, individual afferent  
19 249 fibers that responded to one sweet stimulus typically also responded to several other sweets, and  
20 250 minimally to bitter or sour tastants (Hellekant and Ninomiya 1994; Wang *et al.* 2009). This type of  
21 251 narrow tuning is much less prevalent for taste qualities other than sweet: individual neurons respond  
22 252 quite variably to different salts (Frank *et al.* 2008). However, this feature remains incompletely  
23 253 explored in the periphery as most studies have utilized only limited panels of taste stimuli.

24 254 Whether sensory afferent fibers and their parent ganglion neurons employ patterns of action potentials  
25 255 to encode stimulus identity has been explored to only a limited extent. Different taste stimuli appear to  
26 256 cause primary afferent fibers to fire action potentials with somewhat different patterns, though these  
27 257 differences are not marked (Lawhern *et al.* 2011; Nagai and Ueda 1981; Ogawa *et al.* 1974). Thus,  
28 258 spike discharge pattern may augment and refine the combinatorial coding described above (Nagai and  
29 259 Ueda 1981). Taste coding in the periphery most likely involves activating a combination of afferent  
30 260 fibers having varying tuning capabilities (from specialists to generalists) and subtly different firing  
31 261 patterns. All these factors together play a role in the transmission of information needed to  
32 262 discriminate sweet, sour, salty, bitter, and umami.

33 263 Parenthetically, a key point that should be noted is that to date, recordings from the primary afferent  
34 264 neurons have only been obtained in anesthetized animals. It is possible that some of the distinctions  
35 265 noted below in the response properties of higher level neurons may be attributable to anesthesia.

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3 2664 267 **Hindbrain neurons: evidence for temporal coding**

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7 268 Gustatory afferents from the periphery project directly to the NTS in the brainstem where there is  
8 269 substantial convergence (Whitehead and Frank, 1983; Whitehead, 1986). Cells in the brainstem, NTS  
9 270 and PbN (the main target of projections from the NTS), are generally more broadly tuned than  
10 271 peripheral fibers in both anesthetized (see Spector and Travers 2005, for a review) and awake (see  
11 272 Roussin et al. 2012, but see Nakamura and Norgren 1991) rodents, though there are still groups of  
12 273 neurons in each structure that are narrowly tuned to a single taste quality. Like fibers/cells in the  
13 274 periphery, neurons in the brainstem can become more broadly tuned with changes in stimulus  
14 275 concentration. Moreover, response profiles, defined as the subset of taste qualities that evokes a  
15 276 response, of NTS and PbN cells can change over time (Sammons et al., 2016). This may be due to  
16 277 the changing inputs to these cells as taste receptor cells die and are replaced. Despite such turnover,  
17 278 the network obviously needs to remain stable in its output. It is possible that extensive convergence  
18 279 from neurons with different profiles of sensitivities may support this stability; that is, the loss or addition  
19 280 of a few inputs with different taste sensitivities would have minimal impact on the target cells if there  
20 281 were enough variety in the array of inputs. Further, simultaneous recordings from taste-responsive  
21 282 NTS and PbN cells have shown that NTS with a particular best stimulus are more effective in driving  
22 283 PbN cells with a similar best stimulus, though the same PbN cells receive input from NTS cells with all  
23 284 types of best stimulus preferences (Di Lorenzo and Monroe, 1997; Di Lorenzo et al. 2009). As a  
24 285 changing array of inputs to NTS cells shift their response profiles from one best stimulus to another,  
25 286 simultaneous activation of enough inputs responding to a given best stimulus may also cause PbN  
26 287 cells upstream to shift their best stimulus in kind, as well as modifying the effectiveness of inputs that  
27 288 were activated. Thus, response profiles may change but the overall proportions of the constituents of  
28 289 the network encoding taste stimuli may remain consistent.

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32 290 With a variety of response profiles in the taste-responsive portion of the NTS and PbN, there remains  
33 291 the problem of how confusion among similar-tasting, but not identical, tastants is resolved. As  
34 292 discussed, the across-fiber/neuron patterns may offer one solution, but another might be response  
35 293 dynamics, that is, temporal coding. Variation in the temporal pattern of taste-evoked firing offers a way  
36 294 to disambiguate two tastants that evoke similar response magnitudes within the same cell (Di Lorenzo  
37 295 et al. 2009).

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44 296 Both specialist and generalist neurons have been described in brainstem taste areas in  
45 297 electrophysiological studies with anesthetized animals. Perceptually similar stimuli evoke similar  
46 298 patterns of neuronal population activity, lending support to the combinatorial coding model discussed

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3 299 above (Geran and Travers 2009; Simon *et al.* 2006; Smith *et al.* 2000). However, unlike taste bud cells  
4 300 and sensory afferent neurons, gustatory neurons of the brainstem do exhibit evidence of temporal  
5 301 coding. “Metric space analysis” (MSA; Victor and Purpura 1996; 1997) has been used to quantify this.  
6 302 MSA begins by determining a “distance” between spike trains in terms of the “cost” of making them  
7 303 identical, via adding, deleting, or moving spikes. Adding or removing a spike costs one arbitrary unit.  
8 304 The cost of moving a spike in time by an amount  $t$  is given by  $qt$ , where  $q$  is a parameter that controls  
9 305 the sensitivity of the distance to spike timing. Based on these distances, calculated from repeated  
10 306 neural responses to presentations of several tastants, one can determine two information-theoretic  
11 307 quantities:  $H^{\text{count}}$  and  $H^{\text{spike}}[q]$ .  $H^{\text{count}}$  is the amount of information about taste quality conveyed by spike  
12 308 count alone, and  $H^{\text{spike}}[q]$  is the amount of information about taste quality when spike timing is taken  
13 309 into account.

14 310 In early work using anesthetized rats, spike timing was shown to convey a significant amount of  
15 311 information about taste stimuli in both the Nucleus of the Solitary Tract (NTS; Di Lorenzo and Victor  
16 312 2003) and the Parabrachial Nucleus of the pons (PbN; (Rosen *et al.* 2011), respectively the first and  
17 313 second synapses in the central gustatory pathway in rodents. Specifically, in about half of the taste-  
18 314 responsive cells in NTS (Di Lorenzo and Victor 2003) and PbN (Rosen *et al.* 2011), spike timing  
19 315 contributes to taste quality discrimination – and in both NTS and PbN this contribution was largest in  
20 316 neurons that would appear to be broadly tuned if only spike count were considered. In addition, in the  
21 317 NTS, spike timing contributes significant amounts of information to distinguishing among responses to  
22 318 the components of binary mixtures (Di Lorenzo *et al.* 2009b), between tastants of different  
23 319 concentrations (Chen *et al.* 2011a) and tastants of the same taste quality but different chemical  
24 320 compositions (Roussin *et al.* 2008).

25 321 While evidence for temporal coding of taste stimuli in brainstem structures has been obtained in the  
26 322 anesthetized animal, further studies asked whether there was similar evidence of temporal coding of  
27 323 taste in the alert animal (Roussin *et al.* 2012; Weiss and Di Lorenzo 2012). To that end, rats were  
28 324 implanted with 8-channel microwire electrode bundles aimed at either the NTS or PbN. Following  
29 325 recovery from surgery, mildly water-deprived rats were placed in an experimental chamber with a  
30 326 drinking spout that allowed control of various fluids on a lick-by-lick basis. Taste responses in the NTS  
31 327 and PbN of awake freely licking rats differed in several ways from those recorded under anesthesia.  
32 328 For example, in addition to the typical phasic-tonic time course of response seen under anesthesia,  
33 329 brief lick-by-lick responses were also apparent in many NTS and PbN cells recorded in awake rats. Of  
34 330 these, some cells had responses that progressively increased with successive licks. There were also  
35 331 many cells with very long latency (>2 sec) taste responses that began long after the licks were

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3 332 completed (Roussin *et al.* 2012), which might be the result of stimulation of post-oral receptors during  
4 333 swallowing.

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7 334 Recordings from the NTS (Roussin *et al.* 2012) and PbN (Weiss *et al.* 2014) of awake freely licking  
8 335 rats revealed a rich variety of cell types in addition to those that respond solely to taste. For example,  
9 336 many cells fire in phase with licking, with peak firing rates just at the time of the lick, or between licks.  
11 337 In addition, there are cells that significantly decrease their firing rate during a lick bout. The relative  
13 338 silencing of such cells when the rat engages in consummatory behavior suggests that they may set  
14 339 the initial conditions for the network to acquire sensory information. Moreover, these data underscore  
16 340 the idea that sensory and motor components of gustation are inextricably linked.

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19 341 In a separate series of experiments, the effects of pairing olfactory stimuli with tastants were tested  
20 342 (Escanilla *et al.* 2015). Widespread modulation of taste responses was observed, including changes in  
21 343 response magnitude and latency following taste-odor pairing. MSA of taste- and odor-evoked  
22 344 responses showed that NTS cells were more competent at discriminating tastants when they were  
23 345 presented with odors than when presented alone. This applied for all taste qualities, and whether or  
24 346 not spike timing was taken into account, leading to the hypothesis that brainstem neurons may be  
25 347 most keenly tuned to respond to naturalistic stimuli, that is food, rather than pure chemical exemplars  
26 348 of taste qualities (Escanilla *et al.* 2015). This was tested by presenting complex, natural stimuli such as  
27 349 grape juice (sweet), clam juice (salty), lemon juice (sour) and coffee (bitter). Evoked spike trains in the  
28 350 PbN of awake freely licking rats displayed conveyed significantly more information to naturalistic  
29 351 stimuli than those associated with single compounds (Weiss *et al.* 2014).

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33 352 In conclusion, data from electrophysiological recordings from awake, freely licking rats, underscores  
34 353 the role of the gustatory brainstem as an important node in the neural circuit that controls food  
35 354 identification and ingestion. In addition, dynamics – both intrinsic to the spike trains and related to the  
36 355 lick cycle – are prominent and functionally significant aspects of neural responses.

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39 356 From the gustatory brainstem, afferents target the most medial portion of the ventral posteromedial  
40 357 thalamus. Taste-responsive thalamic neurons in this nucleus form an important source of input to  
41 358 gustatory cortex. Although this small region has been understudied relative to other taste areas, there  
42 359 is recent evidence that the gustatory thalamus may play important roles in taste quality and palatability  
43 360 coding, as well as stimulus expectation (Liu and Fontanini 2015).

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53 362 **Patterns of activity in the rodent gustatory cortex**

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3 363 Within gustatory cortex (GC), physiological studies demonstrate that taste-responsive cells are often  
4 364 multimodal, responding to other sensory stimuli in addition to taste (for review, see Maffei *et al.* 2012).  
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6 365 When recordings are made in either anesthetized or awake animals probed with only sapid stimuli,  
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8 366 both narrowly and broad taste-responsive neurons are found, similar to those found in both peripheral  
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10 367 and other central taste areas (e.g. Katz *et al.* 2001; Ogawa *et al.* 1992a; Ogawa *et al.* 1992b; Sadacca  
11 368 *et al.* 2016; Spector and Travers 2005; Stapleton *et al.* 2006a; Yamamoto *et al.* 1989; Yamamoto *et al.*  
12 369 1984; Yamamoto *et al.* 1985). The roles of these cell types are still ambiguous in terms of function,  
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14 370 although there is evidence that some cortical taste neurons may respond broadly to sets of stimuli that  
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16 371 can be classified as sharing a hedonic value (Fontanini and Katz 2006; Yamamoto *et al.* 1989).

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18 372 An important and related, yet not well-understood aspect of taste coding, involves the spatial  
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20 373 organization of taste neurons – are cells responsive to particular taste stimuli clustered together?  
21 374 Other sensory systems differ in this mode of organization; from the well-known somatotopy of barrel  
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23 375 cortex, to the apparent random overlap of odorant responses in piriform cortex (Petersen 2007;  
24 376 Stettler and Axel 2009). Chen and colleagues (2011b) used 2-photon imaging to describe a sharply  
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26 377 segregated quality representation in mouse GC. Here, quality-specific clusters of singly responsive  
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28 378 neurons were separated in space along the cortical surface, by areas with only sparse taste-evoked  
29 379 activity. In contrast, the vast majority of work on taste cortex is entirely consistent in suggesting that  
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31 380 there is little to no stimulus topography in how taste qualities are represented in the gustatory cortex.  
32 381 Across the anterior – posterior expanse of GC, mapping studies using different techniques have  
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34 382 yielded very different conclusions. For instance, studies using either in vivo recordings, or intrinsic  
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36 383 imaging, show a large degree of overlap among the basic taste stimuli, with bias towards  
37 384 overrepresentation of individual qualities at the anterior and posterior extremes (Accolla *et al.* 2007;  
38 385 Bahar *et al.* 2004; Carleton *et al.* 2010; Yamamoto *et al.* 1985). A genetically encoded trans-synaptic  
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40 386 tracer similarly suggested that neurons receiving input for different taste qualities are intermingled in  
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42 387 the gustatory cortex (Sugita and Shiba 2005).

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44 388 More recently, 2-photon imaging was used to investigate taste responses to stimuli representing four  
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46 389 primary qualities (acid, bitter, salty and sweet) in an area of mouse gustatory cortex defined by taste  
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48 390 thalamic input (Fletcher *et al.* 2017). This “central” area, located just posterior to the landmark middle  
49 391 cerebral artery, possessed thalamic terminal labeling concentrated in the dysgranular subdivision.  
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51 392 Using a virally expressed calcium indicator (GCaMP6s), taste imaging responses were collected in  
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53 393 anesthetized mice in this delineated area. Not surprisingly, cortical taste cells were found to respond  
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55 394 either best to individual stimuli, or combinations of stimuli. Spatial mapping demonstrated that taste  
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57 395 quality responses overlapped in this region, with no evidence of segregation of cells responding to a  
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59 396 single quality. Principle components analysis of this aggregate data suggested that the primary taste

397 qualities were distinctly represented in the population response, providing a basis for discrimination  
398 despite spatial overlap. Moreover, the stimuli were ordered along the first component in a way that  
399 suggested hedonic character may also be represented in the response.

400 The finding of an area of quality overlap in the center of mouse GC fits in nicely with the previously  
401 mentioned mapping studies in the rat (Accolla *et al.* 2007; Yamamoto *et al.* 1985), and other recent 2-  
402 photon approaches in mice (Lavi *et al.* 2018; Livneh *et al.* 2017). Still, these papers and the Chen *et al.*  
403 (2011b) study leave open the possibility that bitter taste responses and sweet taste responses may  
404 be overrepresented posteriorly and anteriorly, respectively, in GC. If so, any topographic  
405 representation of taste quality likely stems from the source of peripheral input. The glossopharyngeal  
406 (IX) nerve, which innervates posterior taste buds, is known to be more responsive to bitter-tasting  
407 stimuli than branches of the facial nerve (VII), which innervate taste buds on the anterior tongue and  
408 palate (Frank 1991; Frank *et al.* 1983). In rat taste cortex, information from the chorda tympani branch  
409 of VII projects to the anterior GC, while information from IX targets the posterior GC (Hanamori *et al.*  
410 1998; Yamamoto *et al.* 1980). A similar “gradient” of taste quality representation that follows peripheral  
411 input has also been described in the parabrachial nucleus in the rodent brainstem (Geran and Travers  
412 2006; Halsell and Travers 1997). In this discussion, however, it is important to consider the multimodal  
413 nature of GC, as well as surrounding cortical areas. For example, there is also a prominent viscerosensory  
414 representation found in posterior insular cortex, adjacent to GC (Cechetto and Saper 1987).  
415 Perhaps correspondingly, the hotspot for conditioned taste aversion learning is also found in posterior  
416 insular or GC (Schier *et al.* 2016; Schier *et al.* 2014). Furthermore, Hanamori and colleagues (1998)  
417 found that over 75% of taste-responsive neurons in posterior GC in rat were also responsive to a  
418 nociceptive stimulus (tail pinch).

419 **In summary, reports (Chen *et al.* 2011b; Peng *et al.* 2015) from a single laboratory notwithstanding,**  
420 **the evidence is now quite strong that gustatory signals for taste quality are distributed and intermingled**  
421 **in the gustatory cortex.**

## 422 **Patterns of gustatory activity in the human cortex**

423 While taste processing in the periphery and also the central nervous system has gained considerable  
424 attention in animal models, these processes are still to be investigated in humans. Of particular  
425 interest are questions on how, when, and where taste information, in general, and specific taste  
426 attributes such as taste quality, intensity, and hedonics, in particular, are processed in the human  
427 brain. Human neuroimaging studies have shown that taste consistently activates a range of cortical  
428 areas including the anterior insula and frontal operculum (FOP), mid-dorsal insula and overlying  
429 Rolandic operculum, posterior insula and POP, as well as the postcentral gyrus (cf. Veldhuizen *et al.*

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3 430 2011; Yeung *et al.* 2018). Evidence suggests that the mid-dorsal insula and the adjacent FOP form  
4 431 GC (Bender *et al.* 2009; Iannilli *et al.* 2012; O'Doherty *et al.* 2001; Small 2010; Small *et al.* 1999), while  
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6 432 the posterior insula and POP have been implicated in oral somatosensation and attention to the mouth  
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8 433 rather than gustation (Veldhuizen *et al.* 2007). These findings are in line with macaque anatomy,  
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10 434 where the anterior and mid insula and the FOP, but not the POP, receive taste afferents from the  
11 435 thalamus (Pritchard *et al.* 1986) but may not directly translate to human physiology. Observations that  
12 436 taste sensations can be elicited by electrical stimulation of the mid-dorsal insula (Mazzola *et al.* 2017)  
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14 437 further corroborate its role as GC. Consistent with the anatomical evidence, scalp-level  
15 438 electrophysiological studies found pronounced activation of the bilateral anterior in mid insula and  
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17 439 adjacent frontal operculum in response to electric (Ohla *et al.* 2010) and sapid taste (Crouzet *et al.*  
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19 440 2015; Tzieropoulos *et al.* 2013) within 150 ms of taste delivery.

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21 441 Functionally, insular activation has been linked with sensory taste features, such as taste intensity  
22 442 (Grabenhorst and Rolls 2008; Guest *et al.* 2007; Ohla *et al.* 2010; Spetter *et al.* 2010; Tzieropoulos *et al.*  
23 443 *et al.* 2013) and taste quality (Crouzet *et al.* 2015; Schoenfeld *et al.* 2004); taste pleasantness and  
24 444 valuation, on the other hand, have been mostly associated with activity in the OFC, the anatomically  
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26 445 later, secondary taste area (Grabenhorst and Rolls 2008; Guest *et al.* 2007). However, it has also  
27 446 been proposed that the GC jointly encodes both the chemical identity and palatability of a tastant (de  
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29 447 Araujo *et al.* 2006) thereby suggesting a role of the insula in the evaluation of taste or its precursors  
30 448 beyond mere sensory processing. This notion is corroborated by observations that expectations about  
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32 449 the value of a taste, induced by visual cues, modulate taste-related processing in the rodent  
33 450 (Grossman *et al.* 2008) and in the human (Nitschke *et al.* 2006; Ohla *et al.* 2012) insula.

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37 451 In contrast to animal models, the mechanisms underlying taste quality coding have received little  
38 452 attention in humans mostly due to the limited spatial resolution of noninvasive brain imaging  
39 453 techniques such as functional magnetic resonance imaging (fMRI) yielding a spatial resolution of a few  
40 454 millimeters at best. Accordingly, only a few fMRI studies have addressed the question of a gustotopic  
41 455 organization of the human GC and their results failed to provide evidence for a clear spatial  
42 456 segregation of taste qualities but rather suggest a partial overlap of insular representations for different  
43 457 tastes (Dalenberg *et al.* 2015; Prinster *et al.* 2017; Schoenfeld *et al.* 2004). However, cortical activation  
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45 458 patterns change rapidly, within milliseconds, rendering temporal information a candidate variable for  
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47 459 taste quality coding. In fact, neuronal response patterns obtained from electrophysiological recordings  
48 460 at the scalp allow deciphering which taste participants tasted on a given trial. The onset of this  
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50 461 discriminability coincided with the earliest taste-evoked responses that were localized in GC signifying  
51 462 that quality is among the first attributes of a taste represented in the central gustatory system (Crouzet  
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53 463 *et al.* 2015) in strong accord with electrophysiological studies in awake rodents (Graham *et al.* 2014;  
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3 464 Pavao *et al.* 2014; Stapleton *et al.* 2006b). The results also align with and add to observations that  
4 465 neuronal response patterns along the rodent gustatory neuroaxis, including the nucleus of the solitary  
5 466 tract (Di Lorenzo *et al.* 2009a), parabrachial nucleus (Geran and Travers 2013), and insula (Jezzini *et*  
6 467 *al.* 2013), code taste quality.

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10 468 More recent evidence linked the predictive value of gustatory neural response patterns and taste-  
11 469 related decision-making. For this, behavioral reports from different tasks were combined with  
12 470 multivariate analyses of large-scale electrophysiological recordings in a series of studies. Specifically,  
13 471 Crouzet and co-workers (2015) showed that the more alike the neural response patterns of any two  
14 472 tastes were, as indicated by poorer discriminative performance of a classifier, the more these tastes  
15 473 were confused by the participants. The results were surprising for the taste domain because they  
16 474 provide evidence for a mapping between neural and phenomenological rather than between neural  
17 475 and chemical spaces. Whether the information encoded in gustatory neural response patterns drives  
18 476 actual behavior was addressed in two further studies. In the study by Wallroth and Ohla (2018),  
19 477 participants were to detect the presence of a taste as quick as possible. They found that the onset of  
20 478 taste decoding (discriminable brain response patterns) indeed predicted *when* participants detected a  
21 479 given taste by button press and linked neuronal response patterns to the speed of simple gustatory  
22 480 perceptual decisions – a vital performance index of nutrient sensing. Interestingly, the onset of taste  
23 481 decoding was earlier in this study, where participants responded speedily, compared to the previous  
24 482 study, where participants performed a delayed response task suggesting that the timing of gustatory  
25 483 coding is in a way flexible and dependent on behavioral goals.

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35 484 While the mere detection of a taste in the oral cavity may prepare a non-specific response, the  
36 485 regulation of nutrient uptake and expulsion of potential toxins calls for quick and reliable taste  
37 486 detection and identification. Whether taste detection and discrimination are sequential or parallel  
38 487 processes, that is whether you know what it is as soon as you taste it, was addressed in another study  
39 488 (Wallroth and Ohla in press). To uncover the sequence of processing steps involved in taste  
40 489 perceptual decisions, participants performed taste-detection and -discrimination tasks. Irrespective of  
41 490 taste quality and task, neural decoding onset and behavioral response times were strongly linked,  
42 491 demonstrating that differences between taste judgments are reflected early during chemosensory  
43 492 encoding. Moreover, neural and behavioral detection times were faster for the iso-hedonic salty and  
44 493 sour tastes than their discrimination time. No such latency difference was observed for sweet and  
45 494 bitter, which differ hedonically. These results indicate that the human gustatory system detects a taste  
46 495 faster than it discriminates between tastes, yet hedonic computations may run in parallel (Perez *et al.*  
47 496 2013) and facilitate taste identification.

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3 497 Together these studies clearly show that the information encoded in taste-related neural response  
4 498 patterns is also the foundation for gustatory decision-making and that the timing aligns with task-  
5 499 specific goals.  
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### 10 501 **Cortical population coding of taste decisions and behavior**

12 502 Taste quality is tightly linked to taste palatability or pleasantness. While sweet taste is typically liked,  
13 503 bitter taste is commonly aversive to most mammals. Accordingly, the gustatory neuroaxis needs to  
14 504 represent both features as they, together, drive food-related decisions and allow adaptive behavior. In  
15 505 awake rats, taste administration is represented by complex temporal coding in single neurons: a brief  
16 506 period of non-specific firing is followed by approximately 500 msec of identity-related firing, which is in  
17 507 turn replaced by firing that is reliably related to taste palatability (Katz *et al.* 2000; Sadacca *et al.*  
18 508 2012). A series of studies have demonstrated that the palatability “epoch” can be independently  
19 509 manipulated, validating the characterization: changes in perceived palatability, such as that observed  
20 510 at the transition from an attentive to “withdrawn” state (Fontanini and Katz 2005; 2006) and across  
21 511 conditioned taste aversion learning (Grossman *et al.* 2008; Moran and Katz 2014), change palatability  
22 512 epoch coding while having no impact on the earlier ~800 ms of taste-induced activity.  
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30 513 CNS neural responses provide information about the identity of tastes on the tongue. Countless  
31 514 studies have demonstrated that sapid stimuli, flowing across the tongue of anesthetized animals,  
32 515 induce responses in neurons across the gustatory neuroaxis (for just a few examples, see (Azuma *et al.*  
33 516 1984; Di Lorenzo 1988; Di Lorenzo and Victor 2003; Erickson *et al.* 1994; Li *et al.* 2013; Nishijo and  
34 517 Norgren 1990; Yamamoto 1984; Yamamoto *et al.* 1989). Perhaps the most discussed facet of these  
35 518 studies is the fact that taste responses vary vastly in breadth; a great deal of energy has been devoted  
36 519 to debating theories of gustatory coding that turn on these breadths of responsivity (e.g., Di Lorenzo  
37 520 2000; Lemon and Katz 2007; Scott 2004; Smith and St John 1999; Spector and Travers 2005). Neural  
38 521 circuitry in general, and taste circuits in particular, are rife with cross-talk and feedback at both micro-  
39 522 (within region) and macro-circuit (between region) level (Jones *et al.* 2006). Empirical and theoretical  
40 523 work makes it clear that neural responses in such interactive networks should contain functionally  
41 524 interpretable dynamics that are most meaningful when examined at the ensemble rather than at the  
42 525 single cell level (e.g., see Abarbanel and Rabinovich 2001).  
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51 526 An independent set of studies have made use of analytic techniques specialized to interpret the real-  
52 527 time firing of multiple simultaneously-recorded neurons (hidden Markov modeling, or HMM). This work  
53 528 reveals that firing rate modulations within taste responses, which appear gradual in across-trial  
54 529 averages of single-neuron firing, are in fact not gradual at all. Rather, they reflect sudden coherent  
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3 530 shifts between ensemble states: at particular time points within individual trials, the firing rates of (on  
4 531 average) ~50% of the recorded neurons will change simultaneously; across-trial averages “smear”  
5 532 these changes, making them look more gradual, because they happen at different latencies in different  
6 533 trials (Jones *et al.* 2007; Miller and Katz 2010). Together, these two sets of results suggest the  
7 534 testable hypothesis that GC neural ensembles, far from simply coding what the taste IS, may process  
8 535 that information to directly drive action. If in fact palatability-related firing appears suddenly in single  
9 536 trials (a possibility implied by but not directly demonstrated in the above-described work), it is possible  
10 537 to hypothesize that this appearance predicts the onset of consumption behavior. Our testing (Sadacca  
11 538 *et al.* 2016) proves this to be the case, in that analyses keyed to the onset of the ensemble state  
12 539 dominant during the palatability epoch (rather than to stimulus onset time, as is more typical) reveal  
13 540 that palatability coding does emerge suddenly—more suddenly than a range of ramping models  
14 541 (including the model used to explain primate perceptual decision-making (see Gold and Shadlen 2001;  
15 542 Shadlen *et al.* 1996) can explain, and as fast as models assuming instantaneous state transitions  
16 543 (Sadacca *et al.* 2016).

17 544 Armed with the knowledge of precisely *when* decision-related information appears in GC on individual  
18 545 trials, the authors were then able to compare this information to within-trial latencies of palatability-  
19 546 related behavioral responses, measured through electromyography. This analysis specifically reveals  
20 547 that the sudden emergence of the “palatability-related state” in GC neural ensembles predicts both  
21 548 *whether* the rat will gape in response to taste stimulation and precisely *when* that gape will occur, in  
22 549 single trials, with correlation values of ~ 0.75 (Sadacca *et al.* 2016).

23 550 The above results, while robust, are phenomenological. Li and co-workers (2016) performed two types  
24 551 of perturbation experiments to test whether GC ensemble transitions are causally linked to  
25 552 consumption behavior. In one experiment, arrival of an aversive taste was cued: as the rats learned  
26 553 the meaning of the cue across a full session, the latency with which they gaped in response to the  
27 554 taste decreased by ~150 ms; recordings showed that the cue had an almost identical impact on neural  
28 555 coding of that aversive taste. In the second experiment; optogenetic silencing of GC neurons was  
29 556 shown to change the likelihood of gaping. Together, these experiments confirm the general hypothesis  
30 557 that GC is a part of a distributed system responsible for transforming an incoming identity code into a  
31 558 taste decision.

32 559 These results, while perhaps surprising within the field of taste research, are consistent with a great  
33 560 deal of work on sensorimotor systems—and, more specifically, on work describing the top-down  
34 561 modulation of multi-rhythmic central pattern generators (Marder 2012).

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## 563 Conclusion

564 When examined at each level of the nervous system – periphery, brainstem, and cortex – it is evident  
565 that individual taste-responsive receptor cells or neurons may respond either selectively or broadly to  
566 stimuli of different taste qualities. Recent approaches to rodent and human central taste also  
567 emphasize the importance of temporal response patterns, which likely underlie the progression of  
568 taste behavior, from detectability to discrimination. This response complexity supports the notion of  
569 combinatorial coding along the gustatory neuroaxis. The flexibility inherent in this type of coding for the  
570 sense of taste may be necessary for animals to exhibit adaptive behavior in food selection and  
571 consummatory behavior.

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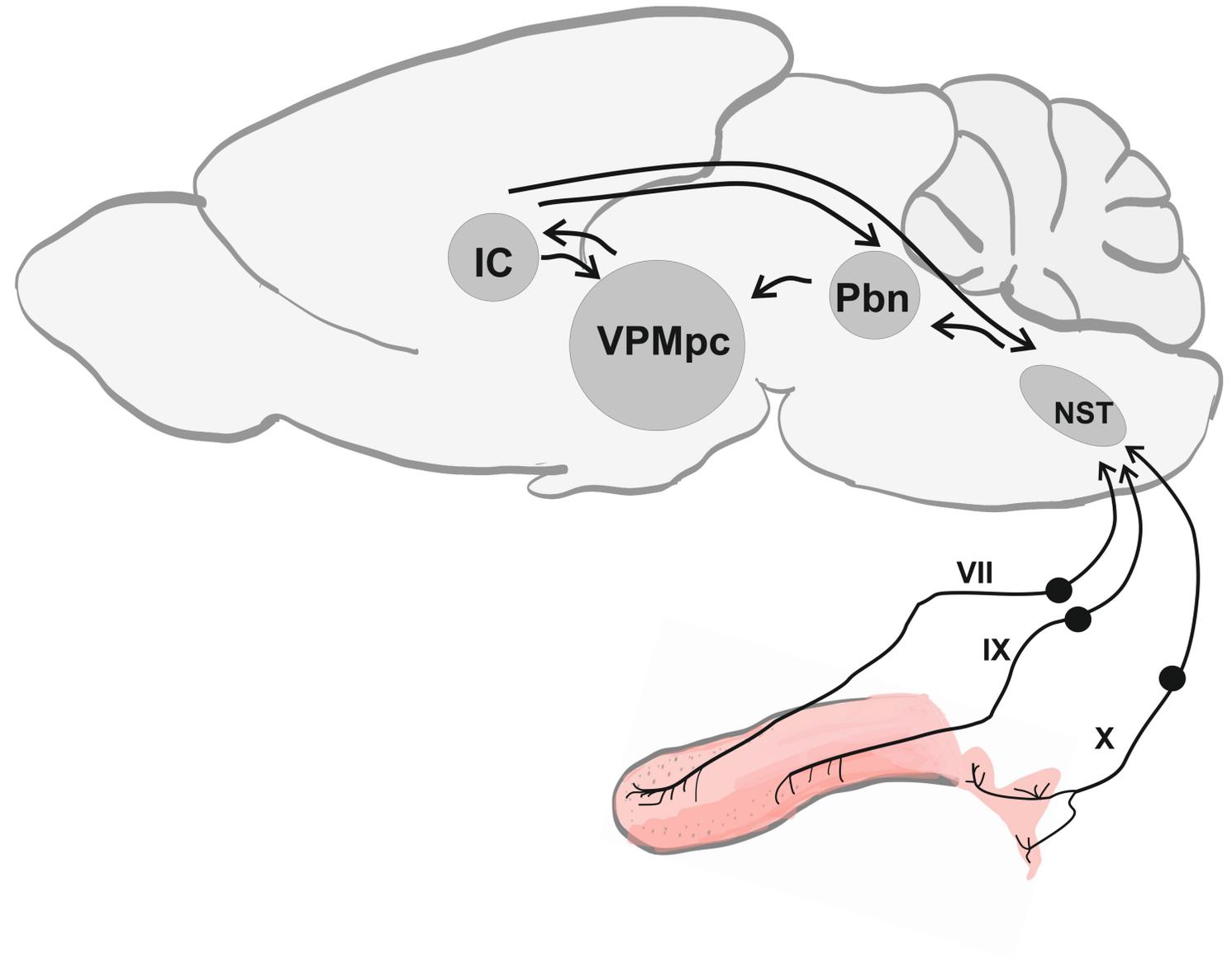
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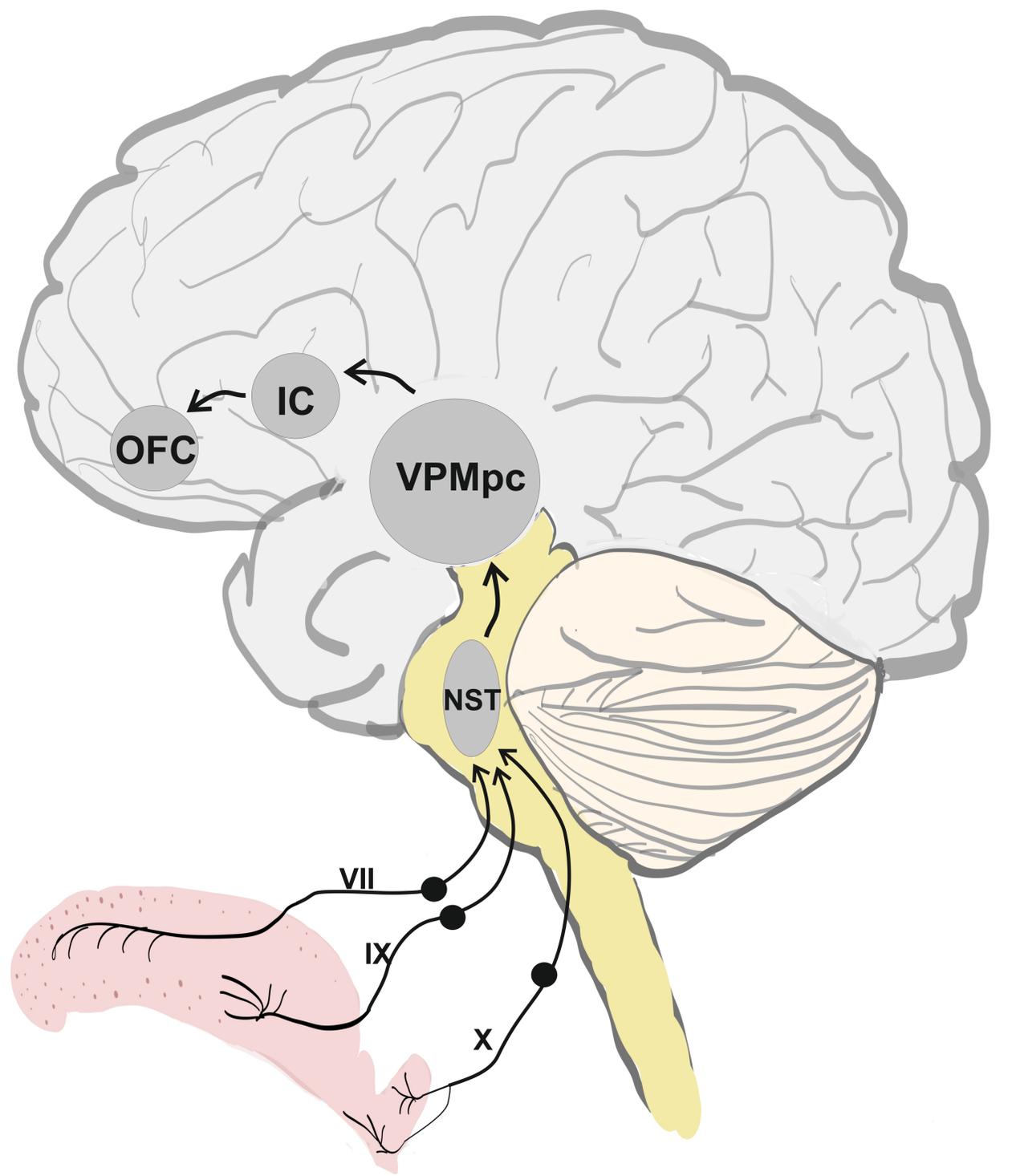
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